

Eff. Date: 14-May-25

Version: 4.1

IFU0020 - DPVB-AP IVDR

# BrightVision, two steps detection system Goat Anti-Mouse/Rabbit IgG AP

#### Instruction For Use

These instructions apply to the WellMed BrightVision; two steps detection system Goat Anti-Mouse/Rabbit AP.

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### 1: Intended use

For in-vitro diagnostic use.

WellMed BrightVision two steps detection system, peroxidase Goat Anti-Mouse/Rabbit IgG AP, is intended for use in immunohistochemistry for the detection of mouse or rabbit antibodies.

### 2: Summary and explanation

The WellMed BrightVision detection system, peroxidase Goat Anti-Mouse/Rabbit AP, is a ready-to-use system that has been manufactured to give an optimal staining, when using the protocol advised in this IFU.

Prior to staining some routine fixed, paraffin-embedded tissue sections should be subjected to pre-treatment (HIER or digestive enzyme).

The BrightVision detection system detects mouse or rabbit antibodies bound to an antigen in tissue sections. The antibodies are not provided but it is recommended to use the WellMed-antibodies. This polymer-complex is then visualized with a suitable substrate/chromogen. The substrate is not provided but it is recommended to use the WellMed-substrate. The clinical interpretation of any staining or its absence should be determined by a qualified pathologist and complemented by morphologic studies; controls should be evaluated within the context of the patient's clinical history and/or other diagnostic tests.

### 3: Kit components

WellMed BrightVision, two steps detection system, peroxidase Goat Anti-Mouse/Rabbit AP.







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## 4: Availability

Catalog Number / Reference	Contents	Volume / Size
DPVB55AP	BrightVision, two steps detection system, Goat Anti-Mouse/Rabbit	2x 55 ml
	AP (ready-to-use)	
	Post-blocking (ready-to-use)	55 ml
	2. Polymer Goat Anti-Mouse/Rabbit AP (ready-to-use)	55 ml
DPVB110AP	BrightVision, two steps detection system, Goat Anti-Mouse/Rabbit	2x 110 ml
	AP (ready-to-use)	
	Post-blocking (ready-to-use)	110 ml
	2. Polymer Goat Anti-Mouse/Rabbit AP (ready-to-use)	110 ml
DPVB500AP	BrightVision, two steps detection system, Goat Anti-Mouse/Rabbit	2x 500 ml
	AP (ready-to-use)	
	Post-blocking (ready-to-use)	500 ml
	2. Polymer Goat Anti-Mouse/Rabbit AP (ready-to-use)	500 ml
DPVB999AP	BrightVision, two steps detection system, Goat Anti-Mouse/Rabbit	2x 1000 ml
	AP (ready-to-use)	
	Post-blocking (ready-to-use)	1000 ml
	2. Polymer Goat Anti-Mouse/Rabbit AP (ready-to-use)	1000 ml

# 5: Recommended staining protocol

Step	Reagent	Template step	Incubation time
1	Deparaffinize and rehydrate tissue section	Slide/tissue preparing	-
2	Wash aqua dest	Wash	2x 5 min
3	If applicable; HIER or digestive enzyme	Pre-treatment	*
4	Wash buffer	TBS buffer	2x 5 min
5	H <sub>2</sub> O <sub>2</sub> (concentration 3%)	Tissue preparing	10 min
6	Wash buffer	TBS buffer	2x 5 min
7	Primary mouse or rabbit antibody	Antibody	30 min
8	Wash buffer	TBS buffer	2x 5 min
9	Detection system, step 1, post-blocking	Post-blocking	15 min
10	Wash buffer	TBS buffer	2x 5 min
11	Detection system, step 2, polymer Mouse/Rabbit AP	Labeled polymer	30 min
12	Wash buffer	TBS buffer	2x 5 min
13	Substrate	Permanent Red	*
14	Wash aqua dest	Wash	2x 2 min
15	Counterstain and coverslip with aqueous mounting medium	Auxiliary	-

<sup>\*</sup> See applicable IFU

### 6: Control slides

A positive control, negative control and reagent control are needed and processed in the same way as the unknown specimen slide to interpret staining results.

# 7: Storage

Store at 2-8 °C and in the dark. Do not use after expiration date.

### 8: Warnings and precautions

Refer to safety data sheet (SDS).







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## 9: Troubleshooting

Please contact WellMed by phone or by email.

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- 1) Shi ZR, Itzkowitz SH, Kim YS. A comparision of three immunoperoxidase techniques for antigen detection in colorectal carcinoma Tissues. J Histochem Cytochem 36: 317-322, 1988
- 2) Loos, C. M. van der. (2007). Multiple Immunoenzyme Staining: Methods and Visualizations for the Observation With Spectral Imaging. Journal of Histochemistry, 56(4), 313–328. doi:10.1369/jhc.2007.950170
- 3) NCCLS. Quality Assurance for Immunocytochemistry; Approved Guideline. NCCLS document MM4-A [1-56238-396-5]. NCCLS, 940 West Valley Road, Suite 1400, Wayne, Pennsylvania 19087-1898 USA, 1999
- 4) Shi ZR, Au A, Soriano R et al: Non-Biotin Amplication (NBA) kit prevents nonspecific background staining of endogenous biotin induced by Heat Induced Epitope Retrieval (HIER) procedure. The J Histotechnol 23:327, 2000
- 5) Mount SL, Cooper K. Beware of biotin: a source of false-positive immunohistochemistry. Current Diagnostic Pathology. 2001; 7:161-167
- Shi SR, Key ME, Kalra KL: Antigen retrieval in formalin-fixed paraffin embedded tissues: An enhanced method for immunohisto-chemical staining based on microwave oven heating of tissue sections. J Histochem Cytochem 39: 741-748, 1991
- 7) Shan-Rong Shi, James Guo, Richard J.cote, Lillian Young, Debra Hawes, Yan Shi, Sandra Thu and Clive R.Taylor, Applied Immunohistochemistry & Molecular Morphology, vol 7,201-208,1999
- 8) Nakane PK and Pierce GB. Enzyme labeled antibodies: Preparations and applications for the localization of antigens. Journal of Histochemistry and Cytochemistry. 1967; 14:929-931
- 9) Battifora, H. Diagnostic Uses of Antibodies to Keratins: A Review and Immunohistochemical Comparison of Seven Monoclonal and Three Polyclonal Antibodies. In: Fenoglio-Preiser, C.M., Wolff, M., Rilke, F. (eds) Progress in Surgical Pathology. Springer, Berlin, Heidelberg. (1988)
- 10) Omata M, Liew CT, Ashcavai M, Peters RL. Nonimmunologic binding of horseradish peroxidase to hepatitis B surface antigen: a possible source of error in immunohistochemistry. American Journal of Clinical Pathology. 1980; 73:626
- 11) Nadji, M., & Morales, A. R. (1983). Immunoperoxidase: Part I. The Technique and Its Pitfalls. Laboratory Medicine, 14(12), 767-771. doi:10.1093/labmed/14.12.767





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